# NUDT15 Variants are Associated with Myelotoxicity and Reduction of 6-Mercaptopurine Dose Intensity in Thai Childhood Acute Lymphoblastic Leukemia การกลายของยืน NUDT15 สัมพันธ์กับภาวะเป็นพิษต่อเซลล์และการลดขนาดยา 6MP ใน โรคมะเร็งเม็ดเลือดขาวชนิดลิมโฟบลาสแบบเฉียบพลันในเด็กไทย

Rawiporn Tiyasirichokchai (รวิพร ติยะศิริโชคชัย)\* Samart Pakakasama (สามารถ ภุคกษมา)\*\* Dr.Chonlaphat Sukasem (คร.ชถภัทร สุขเกษม)\*\*\* Dr.Apichaya Puangpetch (คร.อภิชญา พวงเพีชร์)\*\*\*\*

# ABSTRACT

*NUDT15* variants altered the enzyme activity and caused thiopurine-induced myelotoxicity. Therefore, our study investigated *NUDT15* polymorphisms and association between *NUDT15* and myelotoxic effects in Thai children with acute lymphoblastic leukemia (ALL). Fisher's exact tests indicated that all *NUDT15* variants were significantly associated with neutropenia ( $P = 3.12 \times 10^{-5}$ ) in week 1-8 and thrombocytopenia (P = 0.027) in week 9-24 of maintenance phase. Moreover, the three types of *NUDT15* polymorphisms: wild-type, heterozygous and homozygous variants tolerated 66.67%, 48.8% and 16.67% of 6-mercaptopurine (6MP) protocol dose, respectively. This results concluded that *NUDT15* variants affected 6MP-induced myelotoxicity and dose adjustment should be concerned in each of *NUDT15* diplotypes.

# บทคัดย่อ

การกลาขของยืน NUDT15 มีผลต่อระดับการทำงานของเอนไซม์และทำให้เกิดภาวะเป็นพิษต่อเซลล์ไขกระดูก เมื่อได้รับยากลุ่ม thiopurine ดังนั้นจุดประสงค์ของงานวิจัยคือ การศึกษาความสัมพันธ์ระหว่างการกลายของยืน NUDT15 กับภาวะเป็นพิษต่อเซลล์ในคนไข้เด็กมะเร็งเม็ดเลือดขาวชนิดลิมโฟบลาสแบบเฉียบพลัน ผลการวิจัยนี้พบว่า การกลายของยืน NUDT15 ทุกแบบสัมพันธ์กับภาวะเม็ดเลือดขาวชนิดนิวโตรฟิลต่ำ (*P* = 3.12x10<sup>-5</sup>) ในช่วงสัปดาห์ที่ 1-8 และเกล็ดเลือดต่ำ (*P* = 0.027) ในช่วงสัปดาห์ที่ 9-24 ของการรักษาระยะ maintenance อย่างมีนัยยะสำคัญทางสถิดิ นอกจากนี้คนไข้ที่มียืน NUDT15 แบบปกติ, heterozygous และ homozygous variants สามารถใช้ยา 6MP ได้ 66.67%, 48.8% และ 16.67% ของระดับยามาตรฐานของการรักษา ตามลำดับ ดังนั้นจึงสรุปได้ว่า การกลายของยินมีผลต่อภาวะ เป็นพิษต่อเซลล์เมื่อใช้ยา 6MP และมีผลต่อการปรับลดระดับยาให้เหมาะสมกับการกลายของยิน *NUDT15* แต่ละแบบ

Keywords: *NUDT15* polymorphisms, Myelotoxicity, 6MP คำสำคัญ: ความหลากหลายของยืน *NUDT15* ภาวะเป็นพิษต่อเซลล์ ยา 6MP

<sup>\*</sup> Student, Master of Science Program in Clinical Pathology, Faculty of Medicine, Ramathibodi Hospital, Mahidol University

<sup>\*\*</sup> Associate Professor, Department of Pediatrics, Faculty of Medicine, Ramathibodi Hospital, Mahidol University

<sup>\*\*\*</sup> Associate Professor, Department of Pathology, Faculty of Medicine, Ramathibodi Hospital, Mahidol University

<sup>\*\*\*\*</sup>Assistant Professor, Department of Pathology, Faculty of Medicine, Ramathibodi Hospital, Mahidol University

#### Introduction

6-mercaptopurine (6MP) is the anti-cancer agent which has been used in the maintenance phase of acute lymphoblastic leukemia (ALL) treatment (Timmer et al., 2016). It can improve the event-free survival (EFS) in ALL patient (Relling et al., 1999). 6MP-induced myelotoxicity discontinues the treatment and associates with the opportunistic infections (Relling et al., 1999; Inaba et al., 2017). Non-adherence of treatment could increase the relapse risk (Toyoda et al., 2000). 6MP is a specific substrate of thiopurine methyltransferase (TPMT) that balances the metabolized form of thiopurine drugs (Chouchana et al., 2012). Patient carrying *TPMT* variants could have the higher risk of myelotoxic effects and 6MP dose intolerance (Pogorzelski et al., 2011). Thai population has been found 5% allele frequency of *TPMT\*3C* which presents the decrease enzyme function (Srimartpirom et al., 2004; Mcleod et al., 1999; Szumlanski et al., 1996). However, 6MP-induced myelotoxicity has been observed in ALL patient although *TPMT* is wild-type.

Recently, genome-wide association study (GWAS) reported that Nucleoside diphosphate-linked moiety Xtype motif 15 (*NUDT15*) polymorphism (rs116855232) associated with 6MP dose intensity reduction in ALL patients, and the heterozygous and homozygous variants of T allele indicated the dosage effect (Yang et al., 2015). *NUDT15* is located on the germ line human chromosome and showed the defective function to catalyze the thioguanine triphosphate (TGTP) to thioguanine monophosphate (TGMP) (Moriyama et al., 2016; Valerie et al., 2016). TGTP is the active metabolite that acts as the nucleotide analog to block DNA replication and inhibited the cell proliferation (Fairchild et al., 1986). Moriyama et al. (2016) found that the six diplotypes with the different of the functional levels. *NUDT15\*1* (wild-type) is the normal activity, (compound) heterozygous variants are the intermediate activity and homozygous variants are the low activity.

Chiengthong et al. (2016) presented the association between only *NUDT15:rs116855232* and 6-MP induced neutropenia in ALL children. The medians of cumulative 6-MP dose of patient carrying heterozygous and homozygous variants were lower than those wild-type. Allele frequency of *NUDT15:rs116855232* from this study was 9%. Interestingly, many studies in Asian population reported only rs116855232 minor allele frequency of 10.2%, 10.4%, 7.2% and 12.1% in Japanese, Korean, Indian and Chinese, respectively (Tanaka et al., 2015; Yang et al., 2014; Shah et al., 2017; Zhu et al., 2016) but the other *NUDT15* genotypes have not yet investigated in Thai

#### Objective of this study

Our objectives were to genotype the *NUDT15* in Thai children with ALL, and investigated the association between *NUDT15* variants and myelotoxicity including to compare the 6-MP dose intensity between *NUDT15* wild-type and variants.

# **Materials and Methods**

#### Patients and treatment

One hundred Thai children with ALL were taken part in this retrospective study; the patients were treated at Ramathibodi Hospital, Mahidol University, they received the chemotherapy according to RAMA ALL001 protocol

between 2004- 2017. The duration of treatment for girls was two years and a half, boys was three years and a half. For RAMA ALL001 protocol, the risk groups were classified to three groups: low, standard and high risk groups. For the low risk group, the initial dose of 6-MP and methotrexate (MTX) were 75 mg/m<sup>2</sup>/day daily and 40 mg/m<sup>2</sup> weekly, respectively. Vincristine (VCR) was intravenous 2 mg/m<sup>2</sup> and prednisolone is oral 40 mg/m<sup>2</sup> monthly. Whereas, the standard and high risk group also received intravenous cyclophosphamide 300 mg/m<sup>2</sup> and cytarabine 300 mg/m<sup>2</sup> monthly. The 6-MP dose adjustment based on the CBC and clinical symptoms. 6-MP dose intensity that based on dose adjustment was the ratio between administrated dose and protocol dose (%).

#### **Toxicity definition**

All patients were assessed for the presence of toxicity according to the Common Terminology Criteria for Adverse Events (CTCAE) version 4.0 within the first 6 months of maintenance phase. A patient was classified as having toxicity based on either grade 3 thrombocytopenia or grade 4 neutropenia. Neutropenia (ANC< 500 cells/mm<sup>3</sup>) was defined as grade 4, thrombocytopenia (Plt count< 50,000/ mm<sup>3</sup>) was defined as grade 3. The lowest points of ANC and Plt count were clarified to myelotoxicity. Developing myelotoxicity within the first 8 weeks was termed as early myelotoxicity and week 9-24 was the late myelotoxicity.

#### NUDT15 Genotyping

Genomic DNA (gDNA) was extracted from buffy coat separated by 3ml peripheral blood using Easy Blood Genomic DNA Purification kit according the manufacturer's protocol (GMbiolab, Taichung City, Taiwan). Total gDNA was quantified by Multiskan GO Microplate Spectophotometer (ThermoFisher Scientific, Massachusetts, USA) and prepared to 30 ng/µl.

We genotyped *NUDT15* by Sanger sequencing. Forward and reverse primers for genotyping *NUDT15\*5* (rs186364861, c.52G>A) and *NUDT15\*6* (rs554405994, c.36\_37insGGAGTC) were F1: 5'-GTC ACT TCC TGC CGC TGC CAG-3' and R1: 5'-GCT CAC CCG AAC TCC AGA TGA CC-3'. PCR was performed on CFX96 TouchTM Real-Time PCR detection System (BIORAD, California, USA). The PCR was performed as followed: 95°C for 3 min, followed by 30 cycles of 95°C for 30 sec, 65°C for 30 sec and 72°C for 45 sec, and final extension at 72°C for 10 min. Forward and reverse primers for genotyping *NUDT15\*3* (rs116855232, c.415C>T) and *NUDT15\*4* (rs147390019, c.416G>A) were F2: 5'-GCA AAG CAT CAC TAT GAG TTT-3' and R2: 5'-GCC ACC TAG AGA TGA TTT CCT-3'. The PCR reaction used 57°C of the annealing temperature. The 50 µl PCR was fulfilled by using 1X PCR buffer, 2.6 mM MgCl<sub>2</sub>, 0.4 mM dNTPs, 10pmol of primers, and 1.5 units of Taq DNA polymerase (Invitrogen, Massachusetts, USA). The amplified PCR product of 204 bp from F1R1 and 287 bp from F2R2 were electrophoresed on a 2% agarose gel and selected those PCR products to purify and sequence (ABI3730XL DNA analyzer, ThermoFisher Scientific, USA). *NUDT15* diplotypes were classified according to Moriyama et al. (2016).

# Statistical analysis

Data were analyzed using SPSS version 18. The samples were evaluated for the Hardy-Weinberg equilibrium using Chi-square test. Statistical associations between categorical variables were evaluated using the chi-squared or Fisher's exact tests. The strength of the association was expressed as odd ratios (ORs) with 95% confidence interval (CI) using the genotype, with only the common allele as a reference. Independent Mann-Whitney U-test was compared



the median of dose intensity, ANC and platelet count (Plt) between *NUDT15* wild-type and variants. A two sided test was considered statistically significant with *P*-value less than 0.05.

## Results

# **Characteristic of patients**

One-hundred children with ALL had the median age of 5.9 years and 6-MP mean initial dose was  $58.16\pm20.58$  mg/m<sup>2</sup>/day. ALL patients were *TPMT* wild-type. B-cell ALL patients were the most found in this study as shown in Table 1. The minor allele frequency of rs116855232 was 5% and rs554405994 was 6%. These genotypes distribution were in Hardy- Weinberg equilibrium. The diplotype frequencies of *NUDT15\*1/\*3*, \*1/\*6, \*1/\*2 and \*2/\*2 were 4%, 6%, 4% and 1%, respectively (Table 2).

# Table 1 Characteristics of ALL children on 6-MP therapy

Characteristics	Subject (n=100)
Age (years)	5.9 (1-14)
Female/Male	46/54
Hematologic malignancy types	
- T cell ALL	10
- B cell ALL	88
- Lymphoblastice lymphoma	2
Risk groups	
- Low	42
- Standard	44
- High	14
6-MP initial dose (mg/m <sup>2</sup> /day)	58.16±20.58

# Table 2 Allele and genotype distributions of NUDT15 in ALL children

SNPs	Genotypes	n (100)	Allele frequency (%)	Hardy–Weinberg P-value
rs116855232 (c.415C>T)	CC CT TT	91 8 1	5	0.114
rs554405994 (c.36_37insGGAGTC)	-/- -/ins ins/ins	89 10 1	6	0.257
NUDT15 diplotypes		n (100)		
*1/*1 (wild-type) *1/*3 (rs116855232) *1/*6 (rs554405994) *1/*2(rs116855232,rs554405994) *2/*2		85 4 6 4 1		

# Association between NUDT15 variants and myelotoxicity

We retrospectively analyzed the myelotoxicity in the first 6 months of maintenance phase. The results showed that *NUDT15\*3* and *NUDT15\*6* associated with neutropenia with *P*-value = 0.045 (OR: 10.421; 95% CI: 1.024-106.04) and 0.034 (OR: 6.947; 95% CI: 1.181-40.887) whereas *NUDT15\*2* associated with neutropenia with *P* - value =  $9.67 \times 10^{-4}$  (OR: 37.531; 95% CI: 1.986-708.724) and thrombocytopenia with *P*-value = 0.009 (OR: 16.714; 95% CI: 2.38-117.386) in the early myelotoxicity (Table 3). In the late myelotoxicity, *NUDT15\*6* associated with neutropenia with *P*-value = 0.012 (OR: 15.304; 95% CI: 0.836-280.253) (Table 4). Taken all diplotypes, we found that all *NUDT15* diplotypes were strong associated with neutropenia in the first 8 weeks with *P*-value =  $3.12 \times 10^{-5}$  (OR: 13.895; 95% CI: 3.551-54.363), and thrombocytopenia in week 9-24 with *P*-value = 0.027 (OR: 5.818; 95% CI: 1.354-25.005).

Table 3 Association between NUDT15 diplotypes and myelotoxicity in week 1-8 of maintenance phase

Myelotoxicity (week 1-8)	Yes	No	OR (95% CI)	P-value*		
NUDT15*3:rs116855232						
Neutropenia grade 4 (AN	$IC < 500/ \text{ mm}^3$ )					
NUDT15*1/*3 NUDT15*1/*1	3 (75%) 19 (22.4%)	1 (25%) 66 (77.6%)	10.421 (1.024-106.040)	0.045*		
Thrombocytopenia grade	3 (Plt < 50,000/ mm	n <sup>3</sup> )				
NUDT15*1/*3 NUDT15*1/*1	0 (0%) 7 (8.3%)	4 (100%) 78 (91.7%)	1.163 (0.057-23.743)	1.000		
NUDT15*6:rs554405994	4					
Neutropenia grade 4 (AN	$IC < 500/ \text{ mm}^3$ )					
NUDT15*1/*6 NUDT15*1/*1	4 (66.7%) 19 (22.4%)	2 (33.3%) 66 (77.6%)	6.947 (1.181-40.887)	0.034*		
Thrombocytopenia grade	3 (Plt < 50,000/ mm	n <sup>3</sup> )				
NUDT15*1/*6 NUDT15*1/*1	1 (16.7%) 7 (8.3%)	5 (83.3%) 78 (91.7%)	2.229 (0.228-21.834)	0.434		
NUDT15*2 (rs116855232,rs554405994)						
Neutropenia grade 4 (ANC < 500/ mm <sup>3</sup> )						
NUDT15*1/*2,*2/*2 NUDT15*1/*1	5 (100%) 19 (22.4%)	0 (0%) 66 (77.6%)	37.513 (1.986-708.724)	9.67x10 <sup>-4</sup> *		
Thrombocytopenia grade 3 (Plt < 50,000/ mm <sup>3</sup> )						
NUDT15*1/*2,*2/*2 NUDT15*1/*1	3 (60%) 7 (8.3%)	2 (40%) 78 (91.7%)	16.714 (2.380-117.386	0.009*		
all NUDT15 diplotypes						
Neutropenia grade 4 (AN	$IC < 500/ \text{ mm}^3$ )					
variants wild-type	12 (80%) 19 (22.4%)	3 (20%) 66 (77.6%)	13.895 (3.551-54.363)	3.12x10 <sup>-5</sup> *		
Thrombocytopenia grade 3 (Plt $< 50,000/$ mm <sup>3</sup> )						
variants wild-type	4 (26.7%) 7 (8.3%)	11 (73.3%) 78 (91.7%)	4.052 (1.018-16.125)	0.058		



Myelotoxicity (week 9-24)	Yes	No	OR (95% CI)	P-value*
NUDT15*3:rs1168552	32		·	-
Neutropenia grade 4 (A	$NC < 500/ \text{ mm}^3$ )			
NUDT15*1/*3 NUDT15*1/*1	0 (0%) 39 (45.9%)	4 (100%) 46 (54.1%)	0.131 (0.007-2.505)	0.128
Thrombocytopenia grad	le 3 (Plt < 50,000/	mm <sup>3</sup> )		
NUDT15*1/*3 NUDT15*1/*1	1 (25%) 5 (5.9%)	3 (75%) 80 (94.1%)	5.333 (0.466-61.000)	0.247
NUDT15*6:rs5544059	94			
Neutropenia grade 4 (A	$NC < 500/ \text{ mm}^3$ )			
NUDT15*1/*6 NUDT15*1/*1	6 (100%) 39 (45.9%)	0 (0%) 46 (54.1%)	15.304 (0.836-280.253)	0.012
Thrombocytopenia grad	le 3 (Plt < 50,000/	mm <sup>3</sup> )		
NUDT15*1/*6 NUDT15*1/*1	2 (33.3%) 5 (5.9%)	4 (66.7%) 80 (94.1%)	8.000 (1.170-54.726)	0.066
NUDT15*2 (rs1168552	232,rs554405994)			
Neutropenia grade 4 (A	$NC < 500/ \text{ mm}^3$ )			
NUDT15*1/*2,*2/*2 NUDT15*1/*1	2 (40%) 39 (45.9%)	3 (60%) 46 (54.1%)	0.782 (0.125-4.948)	1.000
Thrombocytopenia grad	le 3 (Plt < 50,000/	mm <sup>3</sup> )		
NUDT15*1/*2,*2/*2 NUDT15*1/*1	1 (20%) 5 (5.9%)	4 (80%) 80 (94.1%)	4.000 (0.374-42.803)	0.298
all NUDT15 diplotype	<b>S</b>			
Neutropenia grade 4 (A	$NC < 500/ \text{ mm}^3$ )			
variants wild-type	8 (53.4%) 39 (45.9%)	7 (46.6%) 46 (54.1%)	1.348 (0.449-4.051)	0.780
Thrombocytopenia grad	le 3 (Plt < 50,000/	mm <sup>3</sup> )		
variants wild-type	4 (26.7%) 5 (5.9%	11 (73.3%) 80 (94.1%)	5.818 (1.354-25.005)	0.027*

Table 4 Association between NUDT15 diplotypes and myelotoxicity at week 9-24 of maintenance phase

# Comparison of ANC, platelet count, dose intensity and NUDT15 polymorphisms

Patients were grouped into two groups according to *NUDT15* wild-type (WT) and variants. *NUDT15* variants (Var) had statistic significantly lower the absolute neutrophil count (ANC) and platelet count (Plt) with *P*-value =  $2.13 \times 10^{-4}$  and 0.009 respectively in week 1- 8 of maintenance phase (Figure 1).

In the first 8 weeks of the maintenance phase, dose intensity of *NUDT* wild-type, heterozygous variants (*NUDT15\*1/\*3*, *NUDT15\*1/\*6* and *NUDT15\*1/\*2*) and homozygous variant (*NUDT15\*2/\*2*) did not statistically significant difference. However, myelotoxicity severally occurred during 8 weeks of maintenance phase then 6MP was adjusted after that on the basis of myelotoxicity and clinical symptoms. The dose reduction was prescribed at the discretion of the treating clinicians. Comparing the dose intensity during week 9-24 of the maintenance phase, the homozygous variant *NUDT15\*2/\*2* had lowest 6MP dose intensity (Figure 2). However, the results showed the trend for the difference of 6MP dose intensity among 3 types of *NUDT15* variants (*P*-value = 0.056).



Figure 1 Box plot of NUDT15 wild-type (WT, n = 85) and variants (Var, n = 15) shows the median of absolute neutrophil count (ANC) (A), and platelet count (Plt) (B) in the week 1-8 of the maintenance phase.

🐨 20<sup>th</sup> NGRC การประชุมวิชาการเสนอผลงานวิจัยระดับบัณฑิตศึกษาแห่งชาติ ครั้งที่ 20 MMO7-8



Figure 2 Comparison of the dose intensity between NUDT15 wild-type (n = 85), heterozygous variants (n = 14) and homozygous variant (n = 1) in the week1-8 and week 9-24. \*P-values were calculated using the Kruskal-Wallis test.

#### Discussion

This was the first report of NUDT15 diplotypes in Thai children with ALL and the results of association between NUDT15 and myelotoxicity, and 6-MP dose intensity. TPMT has been the germ line polymorphism of candidate gene for thiopurine immunosuppressant and anti-cancer drug (Vannaprasaht et al., 2009; Booth et al., 2011). However, myelotoxicity incidence was high in Asians even though the frequency of TPMT polymorphisms was less found compared with Europeans (Kham et al., 2008; Cooper et al., 2008). Our study showed that NUDT15 variants conferred 6MP myelotoxicity and were susceptible to 6MP protocol dose in the patient with TPMT wild-type. According to Yang et al. (2015), they revealed that NUDT15:rs116855232 was the genetic determinant of 6-MP intolerance in ALL children. They observed minor allele frequency of 9.8%, 3.9%, 0.2% and 0% in East Asians Hispanics, Europeans and Africans, respectively. Chiengthong et al. (2016) demonstrated that NUDT15:rs116855232 increased the 6-MP induced myelotoxicity in the maintenance phase of ALL children, and allele frequency of this genotype was 9%. Similarly to Japanese children with ALL, NUDT15:rs116855232 significantly associated the leukopenia (WBC  $\leq 2,000$  cells/mm<sup>3</sup>) during first 2 months of maintenance phase. The allele frequency is 16% of Japanese (Tanaka et al., 2015). Our study detected the minor allele frequency of rs116855232 and rs554405994 were 5% and 6%, respectively. Therefore, the genotyping of rs554405994 can improve the predictive value. Our results showed that NUDT15\*1/\*3 (rs116855232) variant was associated with neutropenia grade 4 in the first 8 weeks of maintenance phase similarly to Chingthong et al. (2016). Furthermore, NUDT15\*1/\*6 (rs554405994) associated with

grade 4 neutropenia in the first 8 weeks in maintenance phase. Whereas, *NUDT15\*1/\*2* (rs116855232, rs554405994) and *NUDT15\*2/\*2* were 100% neutropenia grade 4 and developed thrombocytopenia grade 3 in the first 8 weeks. Especially, a patient carrying *NUDT15\*2/\*2* had pancytopenia with the lowest points of white blood cell count (WBC): 920/mm<sup>3</sup>, platelet (Plt) count: 8,000/mm<sup>3</sup> and 10% Hematocrit before 6-MP dose was adjusted. All association results suggested that genotyping of rs554405994 (c.36\_37insGGAGTC) could be done to avoid thrombocytopenia and neutropenia after 8 weeks of maintenance phase. Thus, the detection of rs554405994 and rs116855232 should be considered for pre-emptive working for 6MP administration in ALL patients.

The median of absolute neutrophil count and platelet count from routine monitoring showed that NUDT15 variants had markedly lower than those wild-type, consistent with Yi et al. (2017). An analysis of dose intensity in week 9-24, NUDT15 wild-type, heterozygous and homozygous diplotypes could be tolerant 6MP 66.67% (50 mg/m<sup>2</sup>/day), 48.8% (36.6 mg/m<sup>2</sup>/day) and 16.67% (12.5 mg/m<sup>2</sup>/day) dose intensity, respectively. Remarkably, even if a patient with NUDT15\*2/\*2 received the lowest dose of all subjects along the maintenance phase; therapy was several interrupted and the myelotoxicity still presented until the treatment ended. It was possible that 6MP 12.5 mg/m<sup>2</sup>/day was not suitable for ALL children with homozygous variant of NUDT15. The tolerant dose in this study was higher than Japanese children with ALL that had the average dose 42.2, 18 and 6 mg/m<sup>2</sup>/day of NUDT15:rs116855232 wildtype, heterozygous and homozygous variants, respectively (Tanaka et al., 2015). While Yang et al. (2015) revealed the significantly diminished 6MP dose intensity in ALL East Asians according to rs116855232 genotype. Their 6MP dose intensity were 75% (56.25 mg/m<sup>2</sup>/day), 47.5% (35.62 mg/m<sup>2</sup>/day) and 10% (7.5 mg/m<sup>2</sup>/day) of NUDT15:rs116855232 wild-type, heterozygous and homozygous variants, respectively. We did not find the association between NUDT15 variants and hepatotoxicity. It could be possible that NUDT15 could metabolize TGTP which plays an important role in the inhibition of cell proliferation then intracellular TGTP accumulation causes the myelosuppression (Valerie et al., 2016). On the other hand, hepatotoxicity correlated with higher 6-methylmercaptopurine ribonucletides (6-MMPR) level (Nygaard et al., 2004). Taken together, NUDT15 variants could develop the sensitivity of TGTP, but may not accumulate 6-MMPR.

Considering the association between *NUDT15* all diplotypes and neutropenia in Table 3, the positive predictive value (PPV) and negative predictive value (NPV) of severe neutropenia was 80% PPV and 77.6% NPV. Thrombocytopenia was predicted with 26.7% PPV and 94.1% NPV, respectively.

#### Conclusion

Our study was the first report the association of *NUDT15* diplotypes with 6MP-induced myelotoxicity and 6MP dose reduction. Furthermore, we developed the genotyping of *NUDT15* polymorphisms and clarify the *NUDT15* diplotypes in Thai children with ALL. *NUDT15* genotyping could be the pre-emptive the 6MP prescription in Thai and other East Asians.

#### Acknowledgement

The authors would like to thank Ramathibodi Cancer Center for the support of reagents, chemicals, clinical data, and specimens. We are grateful to Hematology-Oncology unit in Department of Pediatrics, Pharmacogenomic Laboratory in Department of Pathology for their help with facilitation of laboratory equipment and the good advice of this study.

#### References

- Chiengthong K, Ittiwut C, Muensri S, Sophonphan J, Sosothikul D, Seksan P, et al. *NUDT15* c.415C>T increases risk of 6-mercaptopurine induced myelosuppression during maintenance therapy in children with acute lymphoblastic leukemia. Haematologica. 2016; 101(1): e24-6.
- Chouchana L, Narjoz C, Beaune P, Loriot MA, Roblin X. Review article: the benefits of pharmacogenetics for improving thiopurine therapy in inflammatory bowel disease. Aliment Pharmacol Ther 2012; 35(1): 15-36.
- Cooper SC, Ford LT, Berg JD, Lewis MJ. Ethnic variation of thiopurine S-methyltransferase activity: a large, prospective population study. Pharmacogenomics 2008; 9(3): 303-9.
- Coulthard SA, Howell C, Robson J, Hall AG. The Relationship Between Thiopurine Methyltransferase Activity and Genotype in Blasts From Patients With Acute Leukemia. Blood 1998, 92(8): 2856-62.
- Fairchild CR, Maybaum J, Kennedy KA. Concurrent unilateral chromatid damage and DNA strand breakage in response to 6-thioguanine treatment. Biochem Pharmacol 1986; 35(20): 3533-41.
- Inaba H, Pei D, Wolf J, Howard SC, Hayden RT, Go M, et al. Infection-related complications during treatment for childhood acute lymphoblastic leukemia. Annals of Oncology 2017; 28: 386-392.
- Kham SK, Soh CK, Liu TC, Chan YH, Ariffin H, Tan PL, et al. Thiopurine S-methyltransferase activity in three major Asian populations: a population-based study in Singapore. Eur J Clin Pharmacol 2008; 64(4): 373-9.
- McLeod HL, Pritchard SC, Githang'a J, Indalo A, Ameyaw MM, Powrie RH, et al. Ethnic differences in thiopurine methyltransferase pharmacogenetics: evidence for allele specificity in Caucasian and Kenyan individuals. Pharmacogenetics 1999; 9(6): 773-6.
- Moriyama T, Nishii R, Perez-Andreu V, Yang W, Klussmann FA, Zhao X, et al. *NUDT15* polymorphisms alter thiopurine metabolism and hematopoietic toxicity. Nat Genet 2016; 48(4): 367-73.
- Nygaard U, Toft N, Schmiegelow K. Methylated metabolites of 6-mercaptopurine are associated with hepatotoxicity. Clin Pharmacol Ther 2004; 75(4): 274-81.
- Pogorzelski JP, Rudnicka ET, Kurzawski M, Brodkiewicz A, Adrianowska N, et al. Thiopurine S-Methyltransferase (TPMT) Polymorphisms in Children With Acute Lymphoblastic Leukemia, and the Need for Reduction or Cessation of 6-Mercaptopurine Doses During Maintenance Therapy: The Polish Multicenter Analysis. Pediatr Blood Cancer 2011; 57: 578-582.

- Relling MV, Hancock ML, Boyett JM, Pui CH, Evans WE. Prognostic importance of 6-mercaptopurine dose intensity in acute lymphoblastic leukemia. Blood 1999; 93: 2817–23.
- Shah SA, Paradkar M, Desai D, Ashavaid TF. Nucleoside diphosphate-linked moiety X-type motif 15 C415T variant as a predictor for thiopurine-induced toxicity in Indian patients. J Gastroenterol Hepatol 2017; 32(3): 620-4
- Srimartpirom S, Tassaneeyakul W, Kukongviriyapan V, Tassaneeyakul W. Thiopurine S-methyltransferase genetic polymorphism in the Thai population. Br J Clin Pharmacol 2004; 58(1): 66-70.
- Szumlanski C, Otterness D, Her C, Lee D, Brandriff B, Kelsell D, et al. Thiopurine methyltransferase pharmacogenetics: human gene cloning and characterization of a common polymorphism. DNA Cell Biol 1996; 15(1): 17-30.
- Tanaka Y, Kato M, Hasegawa D, Urayama KY, Nakadate H, Kondoh K, et al. Susceptibility to 6-MP toxicity conferred by a *NUDT15* variant in Japanese children with acute lymphoblastic leukaemia. Br J Haematol 2015; 171(1): 109-15.
- Timmer A, Patton PH, Chande N, McDonald JW, MacDonald JK. Azathioprine and 6-mercaptopurine for maintenance of remission in ulcerative colitis. Cochrane Database Syst Rev. 2016(5): Cd000478.
- Toyoda Y, Manabe A, Tsuchida M, Hanada R, Ikuta K, et al. Six months of maintenance chemotherapy after intensified treatment for acute lymphoblastic leukemia of childhood. J Clin Oncol 2000; 18: 1508-16.
- Valerie NC, Hagenkort A, Page BD, Masuyer G, Rehling D, Carter M, et al. NUDT15 Hydrolyzes 6-Thio-DeoxyGTP to Mediate the Anticancer Efficacy of 6-Thioguanine. Cancer Res 2016; 76(18): 5501-11.
- Vannaprasaht S, Angsuthum S, Avihingsanon Y, Sirivongs D, Pongskul C, Makarawate P, et al. Impact of the heterozygous *TPMT\*1/\*3C* genotype on azathioprine-induced myelosuppression in kidney transplant recipients in Thailand. Clin Ther. 2009; 31(7): 1524-33.
- Yang JJ, Landier W, Yang W, Liu C, Hageman L, Cheng C, et al. Inherited NUDT15 variant is a genetic determinant of mercaptopurine intolerance in children with acute lymphoblastic leukemia. J Clin Oncol 2015; 33(11): 1235-42.
- Yang SK, Hong M, Baek J, Choi H, Zhao W, Jung Y, et al. A common missense variant in *NUDT15* confers susceptibility to thiopurine-induced leukopenia. Nat Genet 2014; 46(9): 1017-20.
- Yi ES, Choi YB, Choi R, Lee NH, Lee JW, Yoo KH, et al. NUDT15 variants cause hematopoietic toxicity with low 6-TGN levels in children with acute lymphoblastic leukemia. Cancer Res Treat 2018; 50(3): 872-882.
- Zhu X, Wang XD, Chao K, Zhi M, Zheng H, Ruan HL, et al. NUDT15 polymorphisms are better than thiopurine Smethyltransferase as predictor of risk for thiopurine-induced leukopenia in Chinese patients with Crohn's disease. Aliment Pharmacol Ther 2016; 44(9): 967-75.