

Chemical Constituents from The Roots of *Dalbergia stipulacea*

## องค์ประกอบทางเคมีจากรากมะขามเฒ่า

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## ABSTRACT

The propose of this study was to extract and purify the chemical constituents from the crude methanol extract of the roots of *Dalbergia stipulacea* by chromatographic methods. The chemical investigation led to the isolation of six known compounds including two pterocarpan derivatives; medicarpin (1) and nitiducarpin (2), two isoflavone derivatives; formononetin (3) and biochanin A (4) and two isoflavane derivatives; nitidulan (5) and dalvelutinane A (6). Their structures were characterized by spectroscopic methods (IR, <sup>1</sup>H-NMR and <sup>13</sup>C-NMR) and by comparison with those of published compounds.

## บทคัดย่อ

การศึกษานี้มีวัตถุประสงค์เพื่อสกัดแยกองค์ประกอบทางเคมีและทำให้บริสุทธิ์จากส่วนสกัดหยาบเมทานอลของรากต้นมะขามเฒ่าด้วยวิธีการทางโครมาโทกราฟี จากการแยกองค์ประกอบทางเคมีพบว่าได้สารประกอบที่มีการรายงานโครงสร้างแล้วทั้งหมด 6 สาร ประกอบด้วย อนุพันธ์เทอโรคาแพน 2 สาร คือ medicarpin (1) และ nitiducarpin (2) อนุพันธ์ของสารไอโซฟลาโวน 2 สาร คือ formononetin (3) และ biochanin A (4) และอนุพันธ์ของสารไอโซฟลาแวน 2 สาร คือ nitidulan (5) และ dalvelutinane A (6) พิสูจน์โครงสร้างของสารเหล่านี้โดยวิธีการทางสเปคโตรสโกปี (IR, <sup>1</sup>H-NMR และ <sup>13</sup>C-NMR) และเปรียบเทียบกับสารที่มีการรายงานโครงสร้างแล้ว

**Keywords:** *Dalberdia stipulacea*, Pterocarpan, Isoflavone

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## Introduction

*Dalbergia stipulacea* (Fabaceae), a climbing shrub, was found in Thailand, Vietnam, Laos, southern China and eastern India. Its roots are used as fish poison (Bhatt et al. 1992). The plant is traditional medicine as infusion of the wood and roots is considered to be emmenagogue. The major compound from the roots of this plant was stipulin (Bhatt et al. 1992). Moreover, luteolin 4'-rutinoside and luteolin were found from its leaves (Borai and Dayal 1993). It was found many flavonoids and isoflavonoids from *Dalbergia* genus such as *D. parviflora* (Songsiang et al. 2009 and 2011; Umehara et al. 2009). Several geranylated flavanones, veluflavanones A-P, were discovered from the stems of *D. velutina* (Kaennakam et al. 2019). Many glycoside derivatives were also isolated from the same plant (Kaennakam et al. 2016). Isoflavones, isoflavone glycosides and neoflavones were isolated from *D. spinosa* (Radha et al. 2015). Moreover, coumarins, chalcones, flavonoids, isoflavonoids glycosides and aurones were found from *D. tonkinensis* (The Son et al. 2018). There is very few studies of chemical substances from *D. stipulacea*. In this study, the crude methanol extract of the roots of *D. stipulacea* was purified and six compounds were isolated.

## Objectives of the study

1. To purify the chemical constituents from the crude methanol extract of the roots of *D. stipulacea* by chromatographic methods.
2. To identify the structures of pure compounds by spectroscopic analysis.

## Materials and methods

### General

The IR spectra were measured with a FTIR Spectrometer: BRUKER TENSOR 27. <sup>1</sup>H and <sup>13</sup>C-NMR spectra were recorded in CDCl<sub>3</sub> using a Varian 400 Plus Spectrometer. Column chromatography (CC) and flash column chromatography (FCC) were carried out on silica gel 60 (0.0063-0.200 and less than 0.0063 mm mesh). Preparative thin layer chromatography was examined on glass-supported silica gel plates using silica gel 60 PF<sub>254</sub> for preparative layer chromatography. Thin layer chromatography (TLC) was carried out on MERCK silica gel 60 F<sub>254</sub> TLC aluminium sheets. TLC spots were visualized by UV light (254 nm) or staining with anisaldehyde reagent.

### Plant material

The roots of *D. stipulacea* were collected in February 2018 from Phuwieng District, Khon Kaen Province, Thailand. The plant was identified by Assoc. Prof. Suppachai Tiaworanant, Faculty of Pharmaceutical Sciences, Khon Kaen University, Thailand. A voucher specimen (KKU012018) was deposited at the Department of Chemistry, Faculty of Science, Khon Kaen University, Thailand.

### Extraction and isolation

The air-dried powdered roots of *D. stipulacea* (11.0 kg) were extracted with hexanes (3×20 L), EtOAc (3×20 L) and MeOH (3×20 L) at room temperature for three days. After evaporation, crude hexanes (162.0 g), EtOAc (368.2 g), and MeOH (375.7 g) extracts were obtained. The crude MeOH was subjected to silica gel CC, eluted with a gradient system of EtOAc:hexanes and EtOAc:MeOH. On the basis of their TLC profile, the fractions which contained the same major compounds were combined to give 11 fractions (MF1-MF11). Fraction MF2 was purified by silica gel CC and eluted with a gradient of 10% EtOAc:hexanes to give nine subfractions, MF2.1-MF2.9. Subfraction MF2.1 was purified on silica gel CC using 2% EtOAc:hexanes as eluent to obtain **1** (27.7 mg). Subfraction MF2.2 was purified on silica gel FCC and 5% acetone:hexanes as eluting solvent to give **2** (12.3 mg). Subfraction MF2.3 was purified by FCC and 5% EtOAc:hexanes as eluting solvent to afford **3** (10.5 mg). Subfraction MF2.4 was separated by FCC and 10% EtOAc:hexanes as eluent to obtain **5** (47.7 mg). Further purification of subfraction MF2.5 by preparative TLC (2% acetone:CH<sub>2</sub>Cl<sub>2</sub>) yielded **6** (6.5 mg). The solid in subfraction MF2.7 was recrystallized (EtOAc:hexanes) to give **4** (3.1 mg).

### Results and discussion

Chromatographic separation of methanol extract of the roots of *D. stipulacea* yielded six known compounds (**1-6**). The structures of all isolated compounds were identified by spectroscopic techniques (IR, <sup>1</sup>H-NMR and <sup>13</sup>C-NMR) and compared with published values. They were (-)-medicarpin (**1**) (Cheng et al. 2019), nitiducarpin (**2**) (Charles and Gandhidasan 2006), formononetin (**3**) (Korbanjhon et al. 2017), biochanin A (**4**) (Songsiang et al. 2009), nitidulan (**5**) (Kaennakam et al. 2017) and dalvelutinane A (**6**) (Kaennakam et al. 2017). (Figure 1).

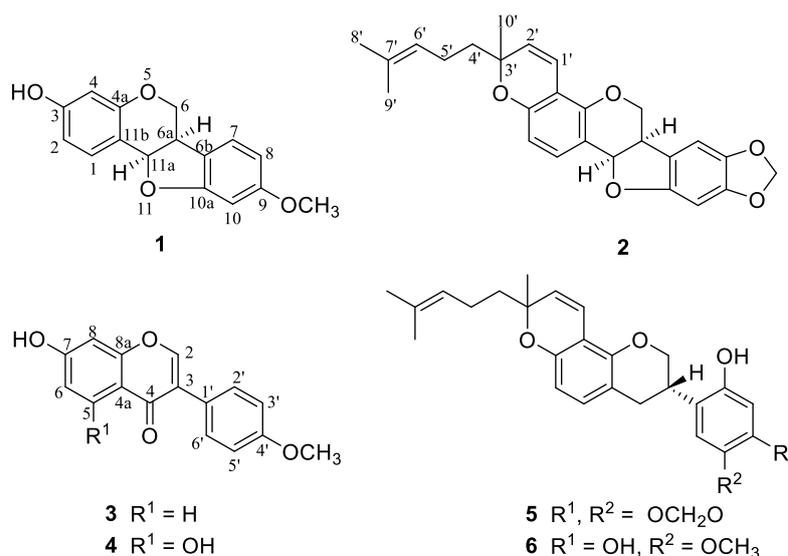


Figure 1 Structures of the isolated compounds from the roots of *D. Stipulacea*

Compound **1** was obtained as a yellowish powder, mp. 123–124 °C (Lit. = 128 °C (Sichaem et al. 2018)). The IR spectrum showed C-H and C-C stretching frequencies of aromatic at 3374 and 1618  $\text{cm}^{-1}$ , respectively. The  $^1\text{H}$  NMR displayed ABX spin system of H-1 ( $\delta_{\text{H}}$  7.30, d,  $J$  = 8.4 Hz), H-2 ( $\delta_{\text{H}}$  6.52, dd,  $J$  = 8.4, 2.4 Hz) and H-4 ( $\delta_{\text{H}}$  6.41, d,  $J$  = 2.4 Hz). The other ABX pattern of H-7 ( $\delta_{\text{H}}$  7.10, d,  $J$  = 8.0 Hz), H-8 ( $\delta_{\text{H}}$  6.36, dd,  $J$  = 8.0, 2.4 Hz) and H-10 ( $\delta_{\text{H}}$  6.39, d,  $J$  = 2.4 Hz) was evident. The  $^1\text{H}$ -NMR signal of H-6 $\alpha$  displayed as doublet of doublets signal at  $\delta_{\text{H}}$  4.17 ( $J$  = 10.8, 4.9 Hz) and H-6 $\beta$  showed triplet signal at  $\delta_{\text{H}}$  3.56 ( $J$  = 10.8 Hz). The signal at  $\delta_{\text{H}}$  5.44 (d,  $J$  = 6.7 Hz) was assigned as H-11a. The methoxyl group exhibited singlet signal at  $\delta_{\text{H}}$  3.72. The  $^{13}\text{C}$ -NMR spectrum contained four oxygenated aromatic carbons at  $\delta_{\text{C}}$  161.0 (C-9), 160.6 (C-10a), 158.4 (C-3) and 156.5 (C-4a). In addition, an oxygenated methylene carbon C-6 exhibited at  $\delta_{\text{C}}$  66.5 while an oxygenated methine carbon C-11a showed at  $\delta_{\text{C}}$  78.8. The comparison of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of **1** corresponded to (-)-medicarpin.

Compound **2** was found as a white amorphous solid, mp. 84-85 °C (Lit. = 84 °C (Heerden et al. 1978)). The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of **2** were similar to those of **1**, except for the presence of an additional geranyl group. Two doublet signals at  $\delta_{\text{H}}$  6.70 ( $J$  = 10.0 Hz, H-1') and 5.54 ( $J$  = 10.0 Hz, H-2') were observed. The olefinic proton H-6' showed triplet signal at  $\delta_{\text{H}}$  5.12 with  $J$  = 6.0 Hz because it was coupled with methylene protons H-5'. Three singlet signals at  $\delta_{\text{H}}$  1.70, 1.60 and 1.41 were assigned as  $\text{CH}_3$ -8',  $\text{CH}_3$ -9' and  $\text{CH}_3$ -10', respectively. In addition, the methylenedioxy group appeared at  $\delta_{\text{H}}$  5.90 and 5.88 and attached to carbon at  $\delta_{\text{C}}$  101.2. The  $^{13}\text{C}$  NMR also showed additional four olefinic carbons at  $\delta_{\text{C}}$  131.5 (C-7'), 128.0 (C-2'), 124.1 (C-6') and 117.0 (C-1'); two methylene carbons at  $\delta_{\text{C}}$  41.1 (C-4') and 22.7 (C-5'); three methyl carbons at  $\delta_{\text{C}}$  26.3 (C-10'), 25.7 (C-8') and 17.6 (C-9'). The oxygenated quaternary carbon at  $\delta_{\text{C}}$  78.8 was determined to C-3'. Thus compound **2** was identified as nitiducarpin.

Compound **3** was obtained as a amorphous solid, mp. 250–251 °C (Lit. = 250–251 °C (Sichaem et al. 2018)). The IR spectrum displayed the presence of carbonyl group at 1599  $\text{cm}^{-1}$ . The  $^{13}\text{C}$  NMR also showed the conjugated carbonyl carbon at  $\delta_{\text{C}}$  174.6 (C-4). Its  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra showed the characteristic signal of isoflavone framework at  $\delta_{\text{H}}/\delta_{\text{C}}$  8.35 (1H, s, H-2)/153.2 (C-2). The ABX pattern of protons on ring A displayed at  $\delta_{\text{H}}$  7.98 (1H, d,  $J$  = 8.7 Hz, H-5), 6.95 (1H, dd,  $J$  = 8.7, 2.1 Hz, H-6) and 6.88 (1H, d,  $J$  = 2.1, H-8). The *para* disubstituted aromatic pattern of ring B showed at  $\delta_{\text{H}}$  7.52 (2H, d,  $J$  = 8.5, H-2' and 6') and 7.00 (2H, d,  $J$  = 8.5, H-3' and 5'). Three oxygenated carbons appeared at  $\delta_{\text{C}}$  162.6 (C-7), 159.0 (C-4') and 157.5 (C-8a). The carbon signals at  $\delta_{\text{C}}$  130.1 (H-2' and H-6') and 113.6 (H-3' and H-5') were evident. A methoxyl group showed signals at  $\delta_{\text{H}}/\delta_{\text{C}}$  3.80 (3H, s)/55.2 in the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra. From all data, compound **3** was determined as formononetin.

Compound **4** was given as a white amorphous solid, mp. 213–214 °C (Lit. = 213–214 °C (Sichaem et al. 2018)). The carbonyl group of this compound showed absorption band at 1650  $\text{cm}^{-1}$  in the IR experiment. In addition, the signal at  $\delta_{\text{C}}$  180.7 of carbonyl carbon was observed in the  $^{13}\text{C}$  NMR data. A 1,4-disubstituted aromatic moiety was evident by showing two doublet signals at  $\delta_{\text{H}}$  7.37 (2H, d,  $J$  = 8.6, H-2' and 6') and  $\delta_{\text{H}}$  6.91

(2H, d,  $J = 8.6$ , H-3' and 5'). Two *meta* coupling protons ( $J = 2.0$  Hz) of H-6 ( $\delta_{\text{H}} 6.25$ ) and H-8 ( $\delta_{\text{H}} 6.29$ ) were detected. The low field signal at  $\delta_{\text{H}} 12.87$  was assigned as intramolecular hydrogen bonding of OH-5. An oxygenated methine carbon at  $\delta_{\text{C}} 152.6$  was identified as C-2, the characteristic signal of isoflavone derivative. The  $^{13}\text{C}$  NMR also exhibited four oxygenated carbons at  $\delta_{\text{C}} 164.0$  (C-7), 162.1 (C-5), 159.6 (C-4'), and 158.2 (C-8a). The high field carbon signals at  $\delta_{\text{C}} 99.2$  and 94.0 were assigned to C-6 and C-8, respectively. The same as **3**, carbons C-2'/C-6' and C-3'/C-5' displayed at  $\delta_{\text{C}} 130.0$  and 114.0, respectively, in the  $^{13}\text{C}$  NMR data. Thus, compound **4** was elucidated as biochanin A.

Compound **5** was obtained as a white amorphous solid, mp. 55-56 °C (Lit. = 55 °C (Heerden et al. 1978)). The IR data displayed O-H stretching band at 3728  $\text{cm}^{-1}$ . In the  $^1\text{H}$  NMR spectrum, two doublet signals ( $J = 8.2$  Hz) at  $\delta_{\text{H}} 6.81$  and 6.36 were assigned as H-5 and H-6, respectively. Two singlet signals at  $\delta_{\text{H}} 6.38$  and 6.60 were assigned to H-3' and H-6', respectively. The prochiral methylene group at C-2 displayed at  $\delta_{\text{H}} 4.34$  (br d,  $J = 10.0$  Hz, H-2a) and 3.99 (t,  $J = 10.0$  Hz, H-2b). In addition, prochiral methylene group at C-4 position showed at  $\delta_{\text{H}} 2.91$  (dd,  $J = 15.0, 10.3$  Hz, H-4a) and 2.80 (dd,  $J = 15.0, 6.7$  Hz, H-4b). The multiplet signal at  $\delta_{\text{H}} 3.50$  (H-3) and broad singlet signal of hydroxy proton at  $\delta_{\text{H}} 4.74$  were found in the  $^1\text{H}$  NMR data. A methylenedioxy group appeared at  $\delta_{\text{H}} 5.89$  and  $\delta_{\text{H}} 5.88$  and were attached on carbon at  $\delta_{\text{C}} 101.1$ . Five oxygenated aromatic carbons at  $\delta_{\text{C}} 152.1$  (C-7), 149.6 (C-8a), 147.8 (C-2'), 146.4 (C-4') and 141.9 (C-5') were determined. The  $\text{sp}^3$  methylene carbons at  $\delta_{\text{C}} 69.9$  (C-2) and 30.6 (C-4) as well as  $\text{sp}^3$  methine carbon at  $\delta_{\text{C}} 31.8$  (C-3) were assigned. This compound contained geranyl moiety were similar to that of **2**. In this part, two doublet signals ( $J = 10.0$  Hz) of H-1'' ( $\delta_{\text{H}} 6.67$ ) and H-2'' ( $\delta_{\text{H}} 5.52$ ) as well as triplet signal ( $J = 7.0$  Hz) of H-6'' were observed. An oxygenated quaternary carbon displayed at  $\delta_{\text{C}} 77.0$  (C-3'') while olefinic carbons appeared at  $\delta_{\text{C}} 124.2$  (C-6'') and 131.5 (C-7'') in the  $^{13}\text{C}$  NMR experiment. Thus, compound **5** was identified as nitidulan.

Compound **6** was found as a yellow viscous oil. The hydroxy group showed the characteristic band at 3389  $\text{cm}^{-1}$  in the IR spectrum. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were similar to those of **5**, except for the disappearance of methylenedioxy group. These spectra showed methoxyl group at  $\delta_{\text{H}}/\delta_{\text{C}} 3.78$  (3H, s)/56.8. Two hydroxy group exhibited as broad singlet at  $\delta_{\text{H}} 5.49$  and 4.62. From all data, compound **6** was determined as dalvelutinane A.

## Conclusions

The chemical investigation of crude methanol extract of *D. stipulacea* roots led to the isolation of six known compounds. From the structural identification by using spectroscopic methods and the comparison with the previous reported, the known compounds were (-)-medicarpin (**1**), nitiducarpin (**2**), formononetin (**3**), biochanin A (**4**), nitidulan (**5**) and dalvelutinane A (**6**).

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